Combinational Effect of *Andrographis paniculata*, *Azadirachta indica* and *Carica papaya* latex for Anthelmintic Potential on *Pontoscolex corethrurus*

Shilpashree V. K.,1 Navesh Velip,2* Deepraj Gaonkar,2 Grishm Kundaikar,2 Chandrakant Gaude,2 Hrithik Raj A. Poosari,2 Ravikumar Nayak3

1 Associate Professor, Department of Pharmacognosy, Karavali College of Pharmacy, Mangaluru, Karnataka, India - 575028.
2 UG Scholar, Department of Pharmacognosy, Karavali College of Pharmacy, Mangaluru, Karnataka, India – 575028.
3 Principal of Karavali College of Pharmacy, Mangaluru, Karnataka, India - 575028

Abstract

The present study was done with the aim to screen the anthelmintic activity of decoction containing *Andrographis paniculata* (Kalmegh), *Azadirachta indica* (Neem), *Carica papaya* latex using *Pontoscolex corethrurus*. The different concentrations (40%, 60% and 80%) of decoction were evaluated for determining the time of paralysis and the time of death of earthworms. Albendazole (10mg/ml) used as standard reference and normal saline as control. The data of the study revealed that the decoction at 80% concentration showed better anthelmintic activity compared to standard drug Albendazole.

2. Materials and methods

2.1. Collection of plant materials

The powders of Kalmegh and Neem were procured from local market of Mangaluru city, Karnataka, India.

2.2. Collection of latex

Unripe matured fruit was tapped early in the morning before making incisions. Vertical cut of 1-2 mm deep was made using a stainless-steel knife. A plastic container was used to collect and to store the latex. Latex adhering to the fruit was carefully scraped off and transferred to the collecting box using plastic spoon. Latex was then subjected for oven drying at 40°C for 45 minutes [11].

2.3. Authentication

The powder samples of Kalmegh and Neem were identified in Department of Pharmacognosy at Karavali college of Pharmacy, Mangaluru, Karnataka.

2.4. Pharmacognostic evaluation

The collected plant materials were studied for organoleptic and powder microscopical characteristics.

2.5. Preparation of decoction

In separate beakers, 25g of each weighed powder sample (Kalmegh and Neem) was thoroughly mixed with 100ml of distilled water and boiled for 15 minutes. The prepared decoction was cooled at room temperature and filtered. The collected plant materials were identified in Department of Pharmacognosy at Karavali college of Pharmacy, Mangaluru, Karnataka.

2.6. Dissolution of latex

10g of dried latex was weighed accurately. Mixed with 50ml of distilled water and filtered. The dissolved latex solution was

10g of dried latex was weighed accurately. Mixed with 50ml of distilled water and filtered. The dissolved latex solution was...
then mixed with the prepared decoction and this was used for further studies.

2.7. Phytochemical screening

Phytochemical analysis was carried out on decoction containing Kalmegh, Neem and Papaya latex for detection of carbohydrates, reducing sugars, proteins and amino acids, alkaloids, glycosides, phenolic compounds, flavonoids, tannins, saponins and phytosterols [12].

Detection of carbohydrates

Molisch's test
To the 2ml of decoction, 2-3 drops of Molisch’s reagent was added followed by 1ml of sulphuric acid along the side of test tube. Formation of violet ring indicates the presence of carbohydrates.

Barfoed’s test
To the 1ml of decoction, 1ml of Barfoed’s reagent was added and heated for 2 minutes. Formation of red precipitate indicates the presence of monosaccharide.

Detection of reducing sugar

Benedict’s test
To 0.5ml of decoction, 0.5ml of Benedict’s reagent was added and boiled for 2 minutes. Appearance of green/yellow colour indicates the presence of reducing sugar.

Fehling's test
1ml of decoction was added to 1ml Fehling’s reagent A & B and boiled in water bath. Formation of red precipitate indicates the presence of reducing sugar.

Detection of Proteins and amino acids

Millon’s test
To the 2ml of decoction add 2ml of Millon’s reagent. Formation of pink coloured solution indicate presence of protein.

Biuret test
To 2ml of decoction add few ml of 2% sodium hydroxide and 10% Copper sulphate solution. Appearance of violet colour indicate presence of amino acid.

Detection of alkaloids

Wagner’s test
To the few ml of decoction, 1-2 drops of Wagner’s reagent was added (along the sides of the test tube). Formation of brown or reddish precipitate indicates the presence of alkaloids.

Picric acid test
To few ml of decoction, 1 drop of 5% picric acid solution was added. Formation of yellow colour indicates the presence of alkaloids.

Detection of glycosides

Concentrated sulphuric acid test
To 5ml of decoction, 2ml of glacial acetic acid and 1 drop of 5% ferric chloride solution was added. To this, add few drops of concentrated sulphuric acid. Formation of blue coloured solution (in acetic acid layer) indicates the presence of glycosides.

Salkowski’s test
To the sample solution add few ml of bromine water. Formation of yellow precipitate indicates presence of cardiac glycoside.

Detection of Phenolic compound:

Lead acetate test
To 1ml of sample solution add few ml of lead acetate solution. Formation of yellow precipitate indicate presence of phenolic compound.

Detection of flavonoids

Shinoda test
To the sample solution add 5ml of alcohol and few magnesium turnings. To this add few drops of concentrated hydrochloric acid. Appearance of pink to crimson colour solution indicate the presence of flavonoids.

Detection of tannins

Ferric chloride test
To aqueous extract add few drops of 5% ferric chloride solution. Appearance of dark green colour indicates the presence of tannins.

Lead subacetate test
To 1ml of decoction add few ml of lead subacetate. Formation of creamy gelatinous precipitate indicates the presence of tannins.

Detection of saponins

Foam test
Take few ml of sample and add few ml of water and shake well. Formation of foam indicates the presence of saponins.

Detection of phytosterol

Salkowski’s test
To the decoction add few ml of concentrated sulphuric acid (shake well and allow to stand). Appearance of red colour in the lower layer indicates the presence of phytosterols [12].

2.8. Anthelmintic screening

Anthelmintic activity was studied for decoction containing Kalmegh powder, Neem powder, and Papaya latex using earthworm species “Pontoscolex Corethrurus”.

2.9. Collection and identification of worms

Adult earthworms (Pontoscolex corethrurus) were collected (due to its anatomical and physiological resemblance with the intestinal parasites) from local garden of Vamanjoor-Mangaluru, Karnataka India. The collected earthworms were identified as Pontoscolex Corethrurus ( Muller 1856) belonging to family Rhinodrilidae. The species were identified by Vivek Hasyagar, Research scholar, Department of applied zoology, Mangaluru university, Konaje-Mangaluru, Karnataka, India.

2.10. Screening method

Five groups of similar sized earthworms were selected. Each group consisted of four earthworms which were used for the study. First group acted as control and received only normal saline. The second group acted as the standard and received Albendazole (10mg/ml). Third, fourth and fifth group served as test and received test sample of 40%, 60% and 80% concentration respectively. The time of paralysis and death of
each individual earthworm was recorded. Paralysis was assumed to occur when the worms were non-motile when introduced into normal saline. The living worms were monitored closely and the time taken for complete death was noted. The death of worms was determined by transferring the motionless worms to warm water at 40°C. The warm environment stimulates and induces movement in the worms, if alive [4, 13].

3. Result & Discussion
The collected plant materials were studied for organoleptic and powder microscopical characteristics. Results are tabulated below

Table 01. Organoleptic evaluation

<table>
<thead>
<tr>
<th>Sl. No</th>
<th>Organoleptic properties</th>
<th>Kalmegh</th>
<th>Neem</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Colour</td>
<td>Pale green</td>
<td>Yellowish green</td>
</tr>
<tr>
<td>2</td>
<td>Odour</td>
<td>Characteristics</td>
<td>Characteristics</td>
</tr>
<tr>
<td>3</td>
<td>Taste</td>
<td>Bitter</td>
<td>Bitter</td>
</tr>
</tbody>
</table>

3.1. Powder microscopy:
The powder microscopy was carried out for powder sample of Andrographis paniculata and Azadirachta indica. Andrographis paniculata shows the presence of epidermis, xylem vessels and elongated fibres tapered at both ends. Azadirachta indica shows the presence of epidermis with anomocytic stomata and covering trichomes.

Table 02. Microscopic evaluation

<table>
<thead>
<tr>
<th>Andrographis paniculata (Kalmegh)</th>
<th>Azadirachta indica (Neem)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Epidermis</td>
<td>Epidermis with anomocytic stomata</td>
</tr>
<tr>
<td>Elongated fibres tapered at both ends</td>
<td>Covering trichome</td>
</tr>
</tbody>
</table>

3.2. Phytochemical analysis:
The phytochemical studies were conducted based on qualitative analysis to identify the presence of bioactive Chemical constituents. Results are tabulated in the table below:

Table 03. Qualitative analysis of phytochemical constituents.

<table>
<thead>
<tr>
<th>Sl. No</th>
<th>Phytochemicals</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Carbohydrates</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>Reducing sugars</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>Proteins &amp; amino acids</td>
<td>-</td>
</tr>
<tr>
<td>4</td>
<td>Alkaloids</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>Glycosides</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>Phenolic compounds</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>Flavanoids</td>
<td>+</td>
</tr>
<tr>
<td>8</td>
<td>Tannins</td>
<td>+</td>
</tr>
<tr>
<td>9</td>
<td>Saponins</td>
<td>+</td>
</tr>
<tr>
<td>10</td>
<td>Phytosterols</td>
<td>+</td>
</tr>
</tbody>
</table>

(+ = Present; (-) = Absent)

Fig. 01. Qualitative tests for phytochemical screening.

3.3. Anthelmintic screening
The herbal preparation containing Andrographis paniculata, Azadirachta indica and Carica papaya latex shows significant anthelmintic activity against Pontoscolex corethrurus. The time of paralysis and time of death of earthworms after being treated with test sample are recorded in table no. 4. The activity of standard drug compared with test sample of different concentrations.

Table 04. Activity of Standard drug compared against test sample at different concentrations.

<table>
<thead>
<tr>
<th>Sr.no</th>
<th>Samples</th>
<th>Concentration</th>
<th>Average time of paralysis (mins)</th>
<th>Average time of death (mins)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Control</td>
<td>Normal saline</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>2</td>
<td>Standard</td>
<td>10mg/ml</td>
<td>162.25</td>
<td>182.00</td>
</tr>
<tr>
<td>3</td>
<td>Test sample</td>
<td>1) 40%</td>
<td>198.25</td>
<td>229.25</td>
</tr>
<tr>
<td>2</td>
<td>Test sample</td>
<td>2) 60%</td>
<td>171.00</td>
<td>194.00</td>
</tr>
<tr>
<td>3</td>
<td>Test sample</td>
<td>3) 80%</td>
<td>127.25</td>
<td>152.5</td>
</tr>
</tbody>
</table>
4. Conclusion
From the above result, it was concluded that the sample prepared from combination of *Andrographis paniculata*, *Azadirachta indica* and *Carica papaya* latex shows significant anthelmintic activity against earthworms *Pontoscolex corethrurus*. It was observed that sample with 80% concentration shows better anthelmintic activity as compared to standard drug Albendazole. So, it was concluded that the combination of these herbs has a good anthelmintic activity against the worms.

5. Acknowledgement
The authors are thankful to Vivek Hasyagar for identification of worms and also thankful to the principal of Karavali College of Pharmacy, Mangaluru, Karnataka, India for providing laboratory facilities to carry out the work.

6. Conflict of Interest
None

7. References